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## Back to the drawing board

### Intestinal neuronal dysplasia type B: not a histological entity yet

Received: 15 October 1997 / Accepted: 12 November 1997

#### Introduction

Intestinal neuronal dysplasia of the submucous plexus (IND B) has been characterised by a group of clinical features indistinguishable from those of Hirschsprung's disease, including constipation, abdominal distension and enterocolitis [28]. The onset may be as early as the first few days of life and it has been commonly associated with Hirschsprung's disease, with which it has been associated with a worse outcome [27, 39]. It has also been associated with pheochromocytoma, medullary carcinoma of the thyroid, carcinoid tumour, cystic fibrosis and paraneoplastic syndromes [8, 29, 40, 41].

The morphological features of IND B were first described by Nezelof et al. in 1970 [29] and clarified further by Meier-Ruge in 1971 [20], naming the condition as neuronc dysplasia of the colon. Continued research led to the description of the pathognomic features of IND B, which included giant ganglia within the submucous plexus [2, 21, 22, 24, 30]. The precise incidence and aetiology of this condition is unknown, but it is cited as a major cause of primary constipation, even commanding its own support group NIDKIDS in Australia [9].

The evidence collected upon this entity is, at first appearances, deceptively infallible. However, there is mounting concern that IND B is still a clinical diagnosis only and not a distinct histopathological entity. There is concern that the previous diagnostic criteria in fact describe the normal large-bowel ganglion cell population which, when recognised in a constipated patient, may even lead to surgical resection. The methodology used to describe the histopathological features of IND B can be readily criticised, and observations and developmental studies suggest that IND B may not exist as a distinct

histopathological entity using the most widely recognised criteria.

#### Methodological criticism

Comparing specimens which have been diagnosed histologically as IND B by virtue of the presence of giant ganglia with normal controls will, of course, prove only that in all the IND B patients giant ganglia are present and that in some of the normal controls they are not [17, 18]. The larger the study, the more significant this result will seem. If this selection bias is unrecognised, there is a danger of concluding that giant ganglia are a diagnostic feature of IND B.

It is also apparent that it is important to first define the normal appearances and quantify normal ranges of the elements to be examined in a study. Several figures for the percentage of ganglia within the submucous plexus of those suffering with the clinical features of IND B have been generated but insufficiencies in the normal control group are all too evident. Kobayashi et al. [13] obtained 10 controls from colostomy sites, but do not reveal what the colostomies were performed for. Similarly, Meier-Ruge et al. [22] obtained 92 normal controls but did not reveal the criteria for taking these biopsies and assessing them as normal. Small numbers of controls and patients also create difficulties in statistical analysis, with less accuracy of significance testing. Kobayashi [12] compared only 23 controls with 9 cases of isolated IND B, and thus difficult significance calculations with Fischer's exact probability test were needed. IND B is currently diagnosed by biopsy of the large bowel. Obtaining a set of controls presents a challenge, as it would be ethically unsound to put a "control" child through an unnecessary surgical biopsy [16]. Koletzko et al. [14] cite this as a major problem with clinicopathological correlation studies in IND B. Meier-Ruge et al. [25] did obtain surgical biopsies from 20 normal control children, up to 15 cm from the pectinate line, and quotes a figure that up to 5% of ganglia are giant ganglia, but does not make it

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clear if this is in normal controls or in patients with IND B. Perhaps the best answer is to use cadaveric material from traumatic death, in which there is no history of constipation and to take into account any possible post-mortem artefact.

Throughout the literature, terms such as “hyperganglinosis” and “hypoganglinosis” are applied freely and frequently and are often recognised as important features of IND B. However, although these words are applied frequently, their use is non-sensical when applied to the submucous plexus: as explained earlier, no-one has defined precisely how many ganglia and ganglion cells there are within the submucous plexus. Smith [37] addresses this problem in the myenteric plexus and accurately describes the counting of ganglion cells within normal and abnormal large bowel, suggesting that hypoganglionosis and hyperganglionosis occur when the number of ganglia are 2 standard deviations below or above the mean, respectively. A similar methodology could be applied to the submucous plexus.

Conveying to the reader the exact method used to perform a scientific study in order that the experiment will be reproducible is one of the cornerstones of all scientific literature. Indeed, in many of the papers examined concerning IND B, complex staining methods and explanations of the statistical tests used were well described and were therefore repeatable. However, the most basic elements within the methods were, on the whole, poorly described. Several sources quantitating ganglia and ganglion cells of the submucous plexus do not define how they identify a cell or whether they included cytoplasmic fragments in the counts or only used whole cells. The fundamental disease process examined, i.e. constipation, is also very poorly described on the whole [21–24, 28]. Loening-Baucke [17] defines constipation as a stool frequency of less than three a week in children. These poor definitions have led to considerable inter- and intra-observer variability. Meier-Ruge in three separate papers produced inconsistent percentages of giant ganglia, although the same methodology was applied throughout [22, 24, 25]. The large variety of methodologies used to describe the neuropathological features of the submucous plexus in constipated children has caused further confusion as to exactly what defines, if anything, IND B (see Lumb and Moore [18] and Schofield and Yunis [35] for details). Lake [16] in a previous editorial has described the confusion that has resulted from constantly changing diagnostic criteria as well as regional differences.

Thus, IND B has grown up around several studies with flawed statistical analysis and poorly defined methodologies with poorly defined and possibly inadequate controls and non-reproducibility of results, and yet it is printed in standard texts of pathology as an entity that a surgeon can treat by cutting out segments of bowel.

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## Developmental study

During development, the submucous nerve plexus develops from primitive neuroblasts derived from the neural crest. Under the influence of homeobox genes, the neuroblasts migrate through the mesenchyme surrounding the developing gut. They are primarily derived from a vagal source and, thus, migration predominantly occurs in a cranio-caudal direction. There is a smaller contribution from a sacral source. The myenteric plexus is sited first, which then gives rise to emigres moving dorsoventrally to form the submucous plexus [5, 11]. The precise mechanism of control of this migration is unknown; whether the migrating blast cells are attracted to their site of eventual maturation or are programmed to migrate to a point [1]. If either of these mechanisms were defective, one would expect that the number of ganglia would be reduced. To produce an area in which there were more ganglia than normal (and thus more giant ganglia), a physical barrier to the neuroblast cell migration would need to be present with continued migratory stimulus. Thus, the neuroblast cells would collect upon the edge of this obstruction and produce a segment with an increased number of giant ganglia, with the proximal portion most effected. Kotiloglu et al. [15] describe an obstructing mucosal web in the small intestine with numerous ganglion cells in the proximal segment. Sacher et al. [32] describe the features of IND B in association with ileal atresia, ileal stenosis and anterior located anus, but also note that it is associated with intussusception, volvulus and meconium peritonitis and propose that IND B is in fact a secondary phenomenon. However, it still remains that with present embryological knowledge, an isolated increase in giant ganglia (that is the feature of IND B) cannot be explained in terms of developmental aberration.

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## Observed findings against IND B

We observed that in resection specimens for rectal carcinoma, the tumour-free resection margins contained giant ganglia which constituted 10% of the total ganglia. Meier-Ruge et al. found that 3–5% [25] and 10% [24] of ganglia were giant in cases of IND B, which suggests that they were merely observing the normal number of giant ganglia. In one study, 56% of a group of adults with primary chronic constipation were diagnosed as having IND B on the basis of the presence of giant ganglia [38]. However, in our experience, ganglia containing more than seven ganglion cells can be found in over half of non constipated adult subjects.

A neurological deficit within the bowel, that causes constipation by rendering the bowel and stool immobile, should have physiological characteristics. However, manometric studies [14, 17, 34, 39] and a radiological study [39] did not find any such pathological physiology.

A disease process and its histological appearance should bear some relationship. Cord-Udy et al. [3] ex-

amined three groups of children, all of whom had presented in early childhood with intestinal dysmobility. Two of the three groups had diagnostic features of IND B on initial biopsy, and one group was histologically normal. There was no significant difference of continued symptomatology after 3–5 years, and absolute numbers of those with continued symptoms were higher in the latter (control) group. Csury and Pena [4] reviewed 25 publications and also concluded that there was no correlation between the histological criteria for IND B and the clinical outcome. Several publications disagree on how common IND B is [6, 19, 26] and this may indicate that this disease has not yet been properly identified or defined.

It is also reported that in those patients diagnosed as suffering with IND B there is an apparent recovery time [26, 39]. But how can this be so if, as originally suggested, IND B is a permanent dysplasia of the control mechanism for propelling a stool through the large intestine? Ryan [31] suggests that there may be some neuronal maturation, but also points out that IND B sufferers are also known to develop progressively worse symptoms. The precise function of the submucous plexus is not known. The most likely function of the most superficial group of ganglion cells (Schabadasch plexus) is secretomotor [33]. Failure of these cells may produce constipation refractory to lubricating laxatives.

### Future research

In order to research entities that cause primary constipation, it is of paramount importance to first understand and characterise the normal features. Most laboratories use haematoxylin and eosin as the baseline stain [16], and any large study into the normal appearance of the enteric nervous system should be first characterised using this stain, detailing the precise staining method and section thickness. Meier-Ruge [21] notes many varying appearances of the innervation of the gut, and describes these as the non-classifiable dysganglinoes. However, which of these non-classifiable dysganglinoes is a normal variant?

Gittes et al. [6] discuss the possibility that IND B may in fact be a disease of unbalanced neuropeptides within the gut and cite vasoactive intestinal peptide and calcitonin-gene related peptide as possible culprits. Kobayashi et al. [13] describe a deficiency in growth-associated protein-43 in those diagnosed with IND B. Hutson et al. [10] describe a deficiency of substance P-reactive fibres in the submucous plexus. Hirobe et al. [7] suggest an immunological mechanism, with an excess of MHC II in the submucous plexus. Ryan [31] details an autoimmune paraneoplastic phenomena in a case of small cell carcinoma of the bronchus.

Recently Shirasawa et al. [36] produced homeobox-deficient mice, deleting the homeobox *Enx* (*Hox11L1*). This homeobox is thought to control the position of the enteric neurones in mice and, when deleted, can produce

similar features to IND B, including a profound constipation, abdominal distension and retarded growth. Histology reveals that these mice, when compared to a normal control group, display an absolute increase in the number of ganglia in the myenteric plexus of the colon but not in the small intestine. Identification of a human homologue for this mouse gene may allow diagnosis of a subgroup of children with chronic constipation circumventing the difficulties of histological diagnosis.

### Conclusions

Intestinal neuronal dysplasia provides a simple explanation of constipation, which easily satisfies the clinician and concerned parents. Up to 95% of constipation in children may in fact be functional [17], and the cure to this common problem does not lie within the jurisdiction of the paediatric gastroenterologist or surgeon. It is important to remember that not all diseases have an underlying histopathological appearance.

The publications reviewed on this subject are all disharmonic and contain inconsistencies and inaccuracies too numerous to enter into a single script. Perhaps, this is the best indication that we need to go back to the drawing board.

### References

- Berry CL (1992) What's in a homeobox. *Virchows Arch* 420:291–294
- Borchard F, Meier-Ruge W, Wiebecke B, Briner J, Münterfering H, Födisch HF, Holschneider A, Schmidt A, Enck P, Stolte M (1991) Innervationsstörungen des Dickdarms. Klassifikation und Diagnostik. *Pathologe* 12:171–174
- Cord-Udy CL, Smith VV, Ahmed S, Risdon RA, Milla PJ (1997) An evaluation of the role of suction rectal biopsy in the diagnosis of intestinal neuronal dysplasia. *J Pediatr Gastroenterol Nutr* 24:1–6; 7–8
- Csury L, Pena A (1995) Intestinal neuronal dysplasia. Myth or reality? Literature review. *Pediatr Surg Int* 10:441–446
- Gershon MD, Epstein ML, Hegstrand L (1980) Colonization of the chick gut by progenitors of enteric serotonergic neurons: distribution, differentiation and maturation within the gut. *Dev Biol* 77:41–51
- Gittes GK, Kim J, Yu G, de Lorimier AA (1993) Severe constipation with diffuse intestinal myenteric hyperganglioneosis. *J Pediatr Surg* 28:1630–1632
- Hirobe S, Doody DP, Ryan DP, Kim SH, Donahoe PK (1992) Ectopic class II major histocompatibility antigens in Hirschsprung's disease and neuronal intestinal dysplasia. *J Pediatr Surg* 27:357–363
- Holschneider A (1982) Hirschsprung's disease. *Hippokrates, Stuttgart*, p 133
- Hutson JM (1996) Intestinal neuronal dysplasia. *Aust Fam Phys* 25:1357
- Hutson JM, Chow CW, Borg J (1996) Intractable constipation with a decrease in substance P-immunoreactive fibres: is it a variant of intestinal neuronal dysplasia? *J Pediatr Surg* 31:580–583
- Kapur R, Yost C, Palmiter RD (1992) A transgenic model for studying development of the enteric nervous system in normal and aganglionic mice. *Development* 116:167–175
- Kobayashi H, Hirakawa H, Puri P (1995) What are the diagnostic criteria for intestinal neuronal dysplasia? *Pediatr Surg Int* 10:459–464

13. Kobayashi H, Hirakawa H, Puri P (1996) Is intestinal neuronal dysplasia a disorder of the neuromuscular junction? *J Pediatr Surg* 31:575–579
14. Koletzko S, Ballauff A, Hadziselimovic F, Enck P (1993) Is histological diagnosis of neuronal intestinal dysplasia related to clinical and manometric findings in constipated children? Results of a pilot study. *J Pediatr Gastroenterol Nutr* 17:59–65
15. Kotiloglu E, Ciftci AO, Tanyel FC, Hicsonmez A (1997) Neuronal intestinal and fibromuscular arterial dysplasia associated with intraluminal mucosal web. *Eur J Pediatr Surg* 7:52–54
16. Lake BD (1995) Intestinal neuronal dysplasia. Why does it only occur in parts of Europe? *Virch Arch* 426:537–539
17. Loening-Bauke V (1994) Constipation in children. *Curr Opin Pediatr* 6:556–561
18. Lumb PD, Moore L (1997) Are giant ganglia a reliable marker of intestinal neuronal dysplasia? *J Pathol* 182 [Suppl]: 33A
19. Martucciello G, Caffarena PE, Lerone M, Mattioli G, Barabino A, Jasonni V (1994) Neuronal intestinal dysplasia: clinical experience in Italian patients. *Eur J Pediatr Surg* 4:287–292
20. Meier-Ruge W (1971) Über ein Erkrankungsbild des colon mit Hirschsprung-Symptomatik. *Verh Dtsch Ges Pathol* 55:506–510
21. Meier-Ruge W (1992) Epidemiology of congenital innervation defects of the distal colon. *Virch Arch* 420:171–177
22. Meier-Ruge W, Gambazzi F, Kaufeler RE, Schmid P, Schmid CP (1994) The neuropathological diagnosis of neuronal intestinal dysplasia (NID B). *Eur J Pediatr Surg* 4:267–273
23. Meier-Ruge WA, Scharli AF, Stoss F (1995) How to improve histopathological results in the biopsy diagnosis of gut dysganglionosis. A methodological review. *Pediatr Surg Int* 10:454–458
24. Meier-Ruge WA, Bronnimann PB, Gambazzi F, Schmid PC, Schmidt CP, Stoss F (1995) Histological criteria for intestinal neuronal dysplasia of the submucous plexus (type B). *Virch Arch* 426:549–556
25. Meier-Ruge W, Schmidt PC, Stoss F (1995c) Intestinal neuronal dysplasia and its morphometric evidences. *Pediatr Surg Int* 10:447–453
26. Milla PJ, Smith VV (1993) Intestinal neuronal dysplasia. *J Pediatr Gastroenterol Nutr* 17:356–357
27. Moore SW, Laing D, Kaschula ROC, Cywes S (1994) A histological grading system for the evaluation of co-existing NID with Hirschsprung's disease. *Eur J Pediatr Surg* 4:293–297
28. Munakata K, Morita K, Okabe I, Sueoka H (1985) Clinical and histological studies of neuronal intestinal dysplasia. *J Pediatr Surg* 20:231–235
29. Nezelof C, Guy-Grand D, Thomine E (1970) Les mégacolons avec hyperplasie des plexus myenteriques. Une entité anatomo-clinique, à propos de 3 cas. *Presse Med* 78:1501–1506
30. Puri P (1997) Variant Hirschsprung's Disease. *J Pediatr Surg* 32:149–157
31. Ryan DP (1995) Neuronal intestinal dysplasia. *Semin Pediatr Surg* 4:22–25
32. Sacher P, Briner J, Hanimann B (1993) Is neuronal intestinal dysplasia (NID) a primary disease or a secondary phenomenon? *Eur J Pediatr Surg* 3:228–230
33. Scheuermann DW, Stach W, Timmermans JP (1991) Functional morphology of the enteric nervous system. *Verh Anat Ges (Anat Anz Suppl)* 170: 85:75–85
34. Schmidt A (1994) Electromanometrical investigations in patients with isolated neuronal intestinal dysplasia (NID). *Eur J Pediatr Surg* 4:310–314
35. Schofield DE, Yunis EJ (1992) What is intestinal neuronal dysplasia? *Pathol Annu* 27: 249–262
36. Shirasawa S, Yunker AMR, Roth KA, Brown GA, Hornig S, Korsmeyer SS (1997) *Enx* (*Hox11L1*)-deficient mice develop myenteric neuronal hyperplasia and megacolon. *Nat Med* 3:646–650
37. Smith VV (1993) Intestinal neuronal density in childhood: a baseline for the objective assessment of hypo- and hyperganglionosis. *Pediatr Pathol Lab Med* 13:225–237
38. Stoss F, Meier-Ruge W (1994) Experience with neuronal intestinal dysplasia (NID) in adults. *Eur J Pediatr Surg* 4:298–302
39. Ure BM, Holschneider AM, Meier-Ruge W (1994) Neuronal intestinal malformations: a retro- and prospective study on 203 patients. *Eur J Pediatr Surg* 4:279–296
40. Whelan T, Gatfield CT, Robertson S, Carpenter B, Schillinger JF (1995) Primary carcinoid of the prostate in conjunction with multiple endocrine neoplasia IIb in a child. *J Urol* 153:1080–1082
41. Wildhaber J, Seelentag WKF, Spiegel R, Schöni MH (1996) Cystic fibrosis associated with neuronal intestinal dysplasia type B. A case report. *J Pediatr Surg* 31:951–954